Volume 7 Issue 2 2021 ISSN 2454-3055

INTERNATIONAL JOURNAL OF ZOOLOGICAL INVESTIGATIONS

Forum for Biological and Environmental Sciences

Published by Saran Publications, India



Alterations in the Prolactin Cells of Stinging Catfish, *Heteropneustes fossilis* Administered with Prolactin and Maintained in Artificial Freshwater or Calcium-Deficient Freshwater

Srivastav Susmita^{1*}, Mishra Diwakar², Srivastav Sunil K.³, Suzuki Nobuo⁴ and Srivastav Ajai K.³

¹Department of Zoology, Shiv Harsh Kisan P. G. College, Basti-272001, India

²Department of Zoology, Government Girls' P. G. College, Ghazipur-233001, India

³Department of Zoology, D.D.U. Gorakhpur University, Gorakhpur-273009, India

⁴Noto Marine Laboratory, Institute of Nature and Environmental Technology, Kanazawa University, Noto-cho, Ishikawa 927-0553, Japan

*Corresponding Author

Received: 25th August, 2021; Accepted: 20th September, 2021; Published online: 27th September, 2021

https://doi.org/10.33745/ijzi.2021.v07i02.045

Abstract: The present study includes the responses of plasma calcium and prolactin cells of a freshwater teleost, *Heteropneustes fossilis* injected with prolactin and maintained in artificial freshwater or calcium-deficient freshwater. Fish were divided into groups A-D. Groups A and B were kept in artificial freshwater. Groups C and D were maintained in calcium-deficient freshwater. Vehicle was administered to groups A and C. Groups B and D were injected with prolactin. Plasma calcium levels and prolactin cells were studied after 1, 3, 5, 10 and 15 days.

In group B, prolactin treatment provoked hypercalcemia from day 3 to day 5; however, the values became normocalcemic at day 10 and day 15.

A significant decrease in the nuclear volume of prolactin cells has been noticed in 5 day prolactin treated fish (group B). This response progresses till day 15. Moreover, on day 10 and day 15, depletion in the cytoplasmic granulation has been observed.

In vehicle-injected fish (group C) the plasma calcium level decreases from day 1 to day 3 (as compared to level of the fish kept in artificial freshwater). Thereafter, the level records an increase from day 5 resulting in hypercalcemia at day 10 and day 15. In prolactin treated fish (group D) the plasma calcium level shows no change up to day 3 as compared to the vehicle-injected group (group C). From day 5 to day 15, the value indicates progressive increase in plasma calcium level.

The prolactin cells of vehicle-injected fish (group C) exhibit hyperactivity on day 3 which is evident by the degranulation, hyperchromaticity of the nuclei and increased nuclear volume. Few cells are seen to degranulate completely after day 5. From day 5 to day 15, the nuclear volume is further increased. Certain cells are seen degenerating on day 10 and day 15.

The prolactin cells of prolactin treated fish (group D) have not shown any change till day 5. Between day 10 and day 15 there is a progressive decrease in the nuclear volume. Moreover, vacuolization and degeneration have also been noticed.

Keywords: Prolactin, Pituitary, Plasma calcium, Stinging catfish, Artificial freshwater, Calcium-deficient freshwater, *Heteropneustes fossilis*

Citation: Srivastav Susmita, Mishra Diwakar, Srivastav Sunil K., Suzuki Nobuo and Srivastav Ajai K.: Alterations in the prolactin cells of stinging catfish, *Heteropneustes fossilis* administered with prolactin and maintained in artificial freshwater or calcium-deficient freshwater. Intern. J. Zool. Invest. 7(2): 615-620, 2021. https://doi.org/10.33745/ijzi.2021.v07i02.001

Introduction

Prolactin (PRL) is a long peptide hormone which is found in all jawed vertebrates (Saha et al., 2021). It is expressed in a variety of different organs and performs diverse physiological functions (Freeman et al., 2000; Srivastav et al., 2017; Dobolyi et al., 2020). In fish, PRL is released into circulation mainly through the definitive mass of prolactin cells present in the pituitary (Brown and Brown, 1987; Srivastav et al., 2017, 2021; Saha et al., 2021). In fishes, prolactin has been associated with many functions - (i) reproduction (Cavaco et al., 2003; Ozaki et al., 2007), (ii) parental behavior (Tacon et al., 2000; Pall et al., 2004; Cunha et al., 2019), (iii) migration (Onuma et al., 2010; Whittington Wilson, and 2013), (iv) immunomodulation (Yada et al., 2004), (v) egg and larval development (Kaneko et al., 2002; Santos et al., 2003), (vi) osmoregulation (Clarke and Bern, 1980; Kelly et al., 1999), and (vii) hypercalcemic action through inhibition of gill calcium efflux (Flik et al., 1994) and by promoting gill calcium influx (Anderson and van Itallie, 2009). In teleosts PRL is designated as "freshwater adopting hormone" (Sanyal and Sen, 2018). Increased calcium accretion in bones and scales of female goldfish has been reported by Takahashi et al. (2008).

With this background an attempt has been made in this study to investigate the effect of administration prolactin in the catfish, maintained either in Heteropneustes fossilis freshwater calcium-deficient artificial or freshwater. The changes induced experimentally by the prolactin in the plasma calcium levels have been correlated with the activity of prolactin cells of pituitary.

Materials and Methods

Live specimens of freshwater catfish *Heteropneustes fossilis* (both sexes) were collected

locally and acclimatized to laboratory conditions for two weeks in plastic pools. For experiments fish were kept in identical glass aquaria each containing 10 L of the medium. 12 fish were kept in each aquarium. The medium was replaced on alternate days. To avoid the effects of circadian rhythm the injections were administered at the same hour of the day (between 8.00 a.m. and 9.00 a.m.). Fish were not fed 24 h before and during the experiment.

Different artificial media i.e. freshwater and calcium-deficient freshwater were prepared as follows;

(a) Artificial freshwater: Distilled water containing (in mmol/liter): NaCl₂.10; Na₂SO₄ 0.45; KCl 0.06; CaCl₂ 0.8; MgCl₂ 0.20. pH of the solution was adjusted to 7.6 with NaHCO₃.

(b) Calcium-deficient freshwater: same as above without CaCl₂.

Adult fish *Heteropneustes fossilis* were divided into 4 numerically equal groups each containing 50 fishes. They were given following treatments:

Group A: Fish were maintained in artificial freshwater and were daily injected intraperitoneally daily with vehicle (0.1 ml of 0.6% NaCl/100 g body wt).

Group B: Fish were kept in artificial freshwater and were daily injected intraperitoneally with 0.1 mg/100 g body wt of Prolactin.

Group C: Fish were maintained in calciumdeficient freshwater and were daily injected intraperitoneally with vehicle.

Group D: Fish were kept in calcium-deficient freshwater and were daily injected intraperitoneally with 0.1 mg/100 g body wt of Prolactin.

Prolactin used in groups B and D was dissolved in 0.6% NaCl solution. Ten fish from each group were anaesthetized with MS 222 and blood samples were taken 2 h after the last injection on 1, 3, 5, 10 and 15 days of the treatment.

Blood samples were collected in heparinized tubes by sectioning of the caudal peduncle. The plasma were separated by centrifugation and analysed for calcium levels (Sigma kits). After collection of blood samples the pituitary gland along with the brain were fixed in aqueous Bouin's fluid and Bouin-Hollande fixative for histological studies. Tissues were routinely processed in graded series of alcohols, cleared in xylene and embedded in paraffin. Serial sections were cut at 6 μ m. The pituitaries were stained with Herlant tetrachrome and Heidenhan's azan technique.

Nuclear (prolactin cells) indexes (maximal length and maximal width) were taken with the aid of ocular micrometer and then nuclear volume was calculated as: volume = $4/3 \pi ab^2$, where 'a' is the major semiaxis and 'b' is the minor semiaxis.

All data were presented as the mean \pm S.E. of six specimens and Student's t test was used to determine statistical significance. In all cases the experimental group was compared to its specific time control group.

Results

(i) Artificial Freshwater (Groups A and B):

No change has been noticed in the plasma calcium levels of vehicle-injected specimens (group A; Fig. 1) throughout the experiment. Following prolactin treatment (group B) the plasma calcium levels remain unaffected on day 1. From day 3 to day 5 the levels progressively increased. The values become normocalcemic at day 10 and day 15 (Fig. 1).

The histological details of the prolactin cells (Fig. 2) of vehicle-injected fish (group A) are more or less similar to those of the details given by Srivastav *et al.* (2017). These cells remain

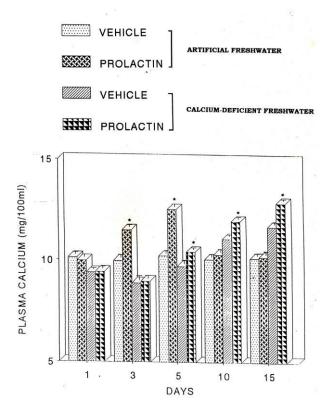


Fig. 1: Plasma calcium levels of *Heteropneustes fossilis* kept either in artificial freshwater or calcium-deficient freshwater and treated with vehicle or prolactin. Each value represents mean \pm S.E. of six specimens. Asterisk indicates significant differences (P<0.05) with vehicle-injected specimens.

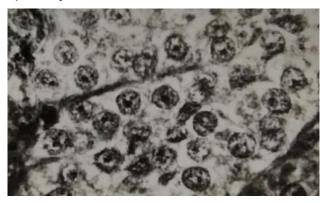


Fig. 2: Prolactin cells of 1 day vehicle-injected *Heteropneustes fossilis* maintained in artificial freshwater. Herlant tetrachrome ×800.

unaltered throughout the experiment.

A significant decrease in the nuclear volume of prolactin cells has been noticed in 5 day prolactin treated fish (group B). This response progresses till day 15 (Fig. 3). Moreover, on day 10 and day 15, a depletion in the cytoplasmic granulation has been observed.

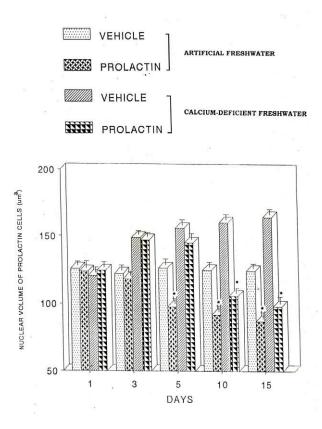


Fig. 3: Nuclear volume of prolactin cells of *Heteropneustes fossilis* kept either in artificial freshwater or calcium-deficient freshwater and treated with vehicle or prolactin. Each value represents mean \pm S.E. of six specimens. Asterisk indicates significant differences (P<0.05) with vehicle-injected specimens.

(ii) Calcium-Deficient Freshwater (Groups C and D):

In vehicle-injected fish (group C) the plasma calcium level decreases from day 1 to day 3 (as compared to level of the fish kept in artificial freshwater). Thereafter, the level records an increase from day 5 resulting in hypercalcemia at day 10 and day 15 (Fig. 1). In prolactin treated fish (group D) the plasma calcium level shows no change up to day 3 as compared to the vehicle-injected group (group C). From day 5 to day 15, the value indicates progressive increase in plasma calcium level (Fig. 1).

The prolactin cells of vehicle-injected fish (group C) exhibit hyperactivity on day 3 which is evident by the degranulation (Fig. 4),

hyperchromaticity of the nuclei and increased nuclear volume (Fig. 1). Few cells are seen to degranulate completely after day 5 (Fig. 5). From day 5 to day 15, the nuclear volume is further increased (Fig. 1). Certain cells are seen degenerating on day 10 and day 15 (Fig. 6).

The prolactin cells of prolactin treated fish (group D) have not shown any change till day 5 (Fig. 1). Between day 10 and day 15 there is a progressive decrease in the nuclear volume (Fig. 1). Moreover, vacuolization and degeneration have also been noticed (Fig. 7).

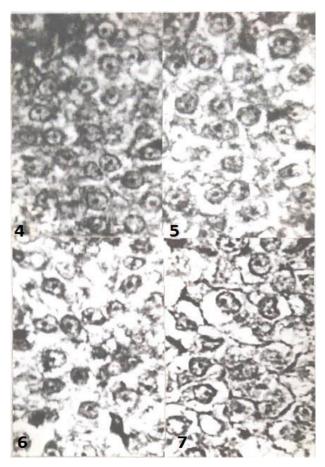


Fig. 4: Degranulation of the prolactin cells of 3 day prolactin injected fish maintained in calcium-deficient freshwater. Herlant tetrachrome x800. Fig. 5: Completely degranulated prolactin cells of 5 day vehiocle-injected fish kept in calcium-deficient freshwater. Herlant tetrachrome x800. Fig. 6: Degeneration in the prolactin cells of 15 day vehicleinjected fish maintained in calcium-deficient freshwater. Herlant tetrachrome x800. Fig. 7: Vacuolization and degeneration in prolactin cells of 15 day prolactin treated fish kept in calcium-deficient freshwater. Herlant tetrachrome x800.

Discussion

In calcium-deficient freshwater the prolactin cells of vehicle-injected *H. fossilis* exhibit degranulation and increased nuclear volume at day 3 and day 5. This can be attributed to the hypocalcemia observed in these specimens. The increased plasma calcium level noticed at day 10 and day 15 seems to be due to the increased release of prolactin from these cells. An enhanced prolactin secretion has been reported in low-calcium freshwater adapted tilapia (Wendelaar Bonga et al., 1985). After acclimation to low-calcium freshwater an increased plasma calcium level has been noticed in tilapia (Wendelaar Bonga et al., 1986). They have stated that this restoration of plasma calcium is most probably mediated by an enhanced production of prolactin hormone.

Prolactin treatment to the fish maintained either in artificial freshwater or calcium-deficient freshwater caused a decrease in the nuclear volume of prolactin cells. This is in conformity with the observations of earlier investigators who have also noticed a decrease in the nuclear volume of prolactin cells after inducing hypercalcemia in the fish by administration of vitamin D₃ and/or CaCl₂ (*Amphipnous cuchia* – Srivastav and Singh, 1989; *Heteropneustes fossilis* – Srivastav *et al.*, 1995). The present study also derives support from the studies of Wendelaar Bonga and Greven (1978) as they have reported that removal of CS causes a three-fold increase in plasma calcium which caused atrophy of prolactin cells.

References

- Anderson JM and Van Itallie CM, (2009) Physiology and function of the tight junction. Cold Spring Harbor Persp Biol. 1(2): a002584.
- Brown PS and Brown SC. (1987) Osmoregulatory actions of prolactin and other adenohypophysial hormones. In: Vertebrate Endocrinology: Fundamentals and Biomedical Implications, (eds.),. Pang, P.K.T., Schreibman, M.P. and Sawyer, W.H., Academic Press, London, pp. 45–84.
- Cavaco JEB, Santos CR, Ingleton PM, Canario AV and Power DM. (2003) Quantification of prolactin (PRL) and PRL receptor messenger RNA in gilthead

seabream (*Sparus aurata*) after treatment with estradiol-17 β . Biol Reprod. 68(2): 588-594.

- Clarke WC and Bern HA. (2012) Comparative endocrinology of prolactin. Horm Proteins Peptides 8: 105-197.
- Cunha AAP, Partridge CG, Knapp R and Neff BD. (2019) Androgen and prolactin manipulation induces changes in aggressive and nurturing behavior in a fish with male parental care. Horm Behav. 116: 104582.
- Dobolyi A, Oláh S, Keller D, Kumari R, Fazekas EA, Csikós V, Renner É and Cservenák M. (2020) Secretion and function of pituitary prolactin in evolutionary perspective. Front Neurosci. 14: 621.
- Flik G, Rentier-Delrue F and Wendelaar Bonga SE. (1994) Calcitropic effects of recombinant prolactins in *Oreochromis mossambicus*. Am J Physiol-Regul Integ Comp Physiol. 266(4): R1302-R1308.
- Freeman ME, Kanyicska B, Lerant A and Nagy G. (2000) Prolactin: structure, function, and regulation of secretion. Physiol Rev. 80(4): 1523-1631.
- Kaneko T, Shiraishi K, Katoh F, Hasegawa S and Hiroi J. (2002) Chloride cells during early life stages of fish and their functional differentiation. Fisheries Sci. 68(1): 1-9.
- Kelly, S.P., Chow, I.N. and Woo, N.Y., 1999. Effects of prolactin and growth hormone on strategies of hypoosmotic adaptation in a marine teleost, Sparus sarba. General and comparative endocrinology, 113(1), pp.9-22.
- Onuma, T.A., Ban, M., Makino, K., Katsumata, H., Hu, W., Ando, H., Fukuwaka, M.A., Azumaya, T. and Urano, A., 2010. Changes in gene expression for GH/PRL/SL family hormones in the pituitaries of homing chum salmon during ocean migration through upstream migration. General and comparative endocrinology, 166(3), pp.537-548.
- Ozaki Y, Ishida K, Saito K, Ura K, Adachi S and Yamauchi K. (2007) Immunohistochemical changes in production of pituitary hormones during artificial maturation of female Japanese eel *Anguilla japonica*. Fisheries Sci. 73(3): 574-584.
- Páll MK. Liljander M and Borg B. (2004) Prolactin diminishes courtship behaviour and stimulates fanning in nesting male three-spined sticklebacks, *Gasterosteus aculeatus*. Behaviour 141(11): 1511-1519.
- Saha I, Chakraborty A and Das S. (2021) Prolactin influences different aspects of fish biology. Asian J Biol Life Sci. 10(1): 51-56.
- Santos CRA, Cavaco JEB, Ingleton PM and Power DM.

(2003) Developmental ontogeny of prolactin and prolactin receptor in the sea bream (*Sparus aurata*). Gen Comp Endocrinol. 132(2): 304-314.

- Sanyal T and Sen K. (2018) Ddiversified role of prolactin in fish: A review. Global J Engineer Sci Res. 5(11): 212-220.
- Srivastav Ajai K and Singh P. (1989) Response of prolactin cells of the freshwater mud eel, *Amphipnous cuchia* to vitamin D_3 administration. Zool Jb Physiol. 93: 235-240.
- Srivastav Ajai K, Singh S and Sasayama Y. (1995) Vitamin D_3 induced changes in the prolactin cells of the fish, *Heteropneustes fossilis* reared in artificial freshwater, calcium-rich freshwater or calciumdeficient freshwater. J Reproduct Biol Comp Endocrinol. 7: 72-82.
- Srivastav S, Mishra D, Srivastav SK, Suzuki N, Srivastav Ajai K. (2017) Estradiol affects prolactin producing cells and calcium levels in a teleost, *Heteropneustes fossilis,* kept in different calcium concentrations. Iranian J Toxicol. 11(5): 45-51.
- Srivastav S, Mishra D, Kumar A, Srivastav SK, Suzuki N and Srivastav Ajai K. (2021) Prolactin induced cytoarchitectural changes in the corpuscles of Stannius of stinging catfish, *Heteropneustes fossilis* acclimated to different calcium media. Intern J Biol Environ Invest. 1 (1): 1-9.
- Tacon P, Baroiller JF, Le Bail PY, Prunet P and Jalabert B. (2000) Effect of egg deprivation on sex steroids, gonadotropin, prolactin, and growth hormone profiles during the reproductive cycle of the mouthbrooding cichlid fish *Oreochromis niloticus*. Gen Comp Endocrinol. 117(1): 54-65.

- Takahashi H, Suzuki N, Takagi C, Ikegame M, Yamamoto T, Takahashi A, Moriyama S, Hattori A and Sakamoto T. (2008) Prolactin inhibits osteoclastic activity in the goldfish scale: a novel direct action of prolactin in teleosts. Zool Sci. 25(7): 739-745.
- Wendelaar Bonga SE and Greven JAA. (1978) The relationship between prolactin cell activity, environmental calcium and plasma calcium in the teleost *Gasterosteus aculeatus*. Observations on stanniectomized fish. Gen Comp Endocrinol. 36: 90-101.
- Wendelaar Bonga SE, Flik G, Lowik CW, Van Eys CJ. (1985) Environmental control of prolactin secretion in the teleost fish *Oreochromis* (formerly *Sarotherodon*) *mossambicus*. Gen Comp Endocrinol. 57(3): 352-359.
- Whittington CM and Wilson AB. (2013) The role of prolactin in fish reproduction. Gen Comp Endocrinol. 191: 123-136.
- Yada T, Misumi I, Muto K, Azuma T and Schreck CB. (2004) Effects of prolactin and growth hormone on proliferation and survival of cultured trout leucocytes. Gen Comp Endocrinol. 136(2): 298-306.